Journal: The American Journal of Clinical Nutrition

Article doi: 10.1093/ajcn/ngy058

Article title: Plasma trimethylamine-N-oxide and related metabolites are associated with type 2

diabetes risk in the Prevención con Dieta Mediterránea (PREDIMED) trial

First Author: Christopher Papandreou Corr. Author: Christopher Papandreou



We encourage you to use Adobe's editing tools (please see the next page for instructions). If this is not possible, please list clearly in an e-mail. Please do not send corrections as track changed Word documents.

Changes should be corrections of typographical errors only. Changes that contradict journal style will not be made.

These proofs are for checking purposes only. They should not be considered as final publication format. The proof must not be used for any other purpose. In particular we request that you: do not post them on your personal/institutional web site, and do not print and distribute multiple copies. Neither excerpts nor all of the article should be included in other publications written or edited by yourself until the final version has been published and the full citation details are available. You will be sent these when the article is published.

- 1. **Licence to Publish:** Oxford Journals requires your agreement before publishing your article. If you haven't already completed this, please sign in with your My Account information and complete the online licence form. Details on how to do this can be found in the Welcome to Oxford Journals email.
- Permissions: Permission to reproduce any third party material in your paper should have been obtained prior to acceptance. If your paper contains figures or text that require permission to reproduce, please inform me immediately by email.
- 3. **Author groups:** Please check that all names have been spelled correctly and appear in the correct order. Please also check that all initials are present. Please check that the author surnames (family name) have been correctly identified by a pink background. If this is incorrect, please identify the full surname of the relevant authors. Occasionally, the distinction between surnames and forenames can be ambiguous, and this is to ensure that the authors' full surnames and forenames are tagged correctly, for accurate indexing online.
- 4. **Figures:** If applicable, figures have been placed as close as possible to their first citation. Please check that they are complete and that the correct figure legend is present. Figures in the proof are low resolution versions that will be replaced with high resolution versions when the journal is printed.
- 5. Missing elements: Please check that the text is complete and that all figures, tables and their legends are included.
- 6. **Special characters and equations:** Please check that special characters, equations and units have been reproduced accurately.
- 7. URLs: Please check that all web addresses cited in the text, footnotes and reference list are up-to-date.
- 8. **Funding:** If applicable, any funding used while completing this work should be highlighted in the Acknowledgements section. Please ensure that you use the full official name of the funding body.



AUTHOR QUERIES - TO BE ANSWERED BY THE CORRESPONDING AUTHOR

The following queries have arisen during the typesetting of your manuscript. Please answer these queries by marking the required corrections at the appropriate point in the text.

| Query No. | Nature of Query | Author's Response |
|-----------|--|--|
| Q1 | Author: Please check title as edited per Journal style. | |
| Q2 | Author: Disclosures: Please ensure that this statement is complete and includes any necessary disclosures | |
| Q3 | Author: Verify accuracy of author names and affiliation pairings. Note that middle initials for Georgios A. Fragkiadakis and Frank B Hu were retained per the manuscript version. | |
| Q4 | Author: Please spell out abbreviations in Affiliations if not already done. Please also verify that affiliations are numbered according to each individual department/division. If multiple divisions within an institution are labeled with one superscript number, please amend to use separate (consecutive) numbers for each division. | \bar{\bar{\bar{\bar{\bar{\bar{\bar{ |
| Q5 | Author: Journal style is to use the systematic name (e.g., C18:0) at first mention and then use the common name (e.g., stearic acid) thereafter. If possible, please amend accordingly throughout. | |
| Q6 | Author: Please check keywords as set. | |
| Q7 | Author: "such as" correct as edited? Or please clarify. | |
| Q8 | Author: Author "MAM-G" is not one of the current authors. Please correct. | |
| Q9 | Author: Please confirm or amend Support footnote as edited. | |
| Q10 | Author: Do you mean "and at the 1-y follow-up" instead of "and yearly during the follow-up"? Please check wording. | |
| Q11 | Author: Please confirm or amend edits for wording in sentence beginning "Insulin resistance was estimated by" | |
| Q12 | Author: "at the 1-y follow-up" ok as edited? | |
| Q13 | Author: "mean \pm SD BMI" correct as edited? | |
| Q14 | Author: Please clarify what the "a:" and "b:" represent here? | |
| Q15 | Author: Please reword "plasmalogen, either metabolites were modeled" to clarify. | |
| Q16 | Author: Note that table layouts/footnotes were edited to better conform to Journal style. Please check carefully. | |
| Q17 | Author: Table 3: Should "MV" be specified as MV1 or MV2? | |
| Q18 | Author: Tables 2 and 3 footnotes: "Values are HRs (95% CIs)" correct as added? Or please amend. | |
| Q19 | Author: Please check figure legends as edited. | |
| Q20 | Author: Is there a word(s) missing after "continuous" in Figure 1 legend? | |
| Q21 | Author: "FDR-controlled" ok as edited? Check similar instances in tables. | |
| Q22 | Author: "and T2D" correct as edited? | |
| Q23 | Author: Fig 2 legend: Note that ": biomarker, or bystander, in T2D?" was deleted? Or if needed, please reword to clarify. | |
| Q24 | Author: "and at 1 y, respectively" correct as edited? | |

| Query No. | Nature of Query | Author's Response |
|-----------|---|-------------------|
| Q25 | Author: "oxygenase or reductase" ok as edited to avoid use of virgule "/" between words? | |
| Q26 | Author: "which was significantly correlated" ok as edited? Or please clarify wording. | |
| Q27 | Author: Please add expansion of GLUT4. | |
| Q28 | Author: Please check author responsibilities section as edited and add specific responsibilities for authors EY, LL, GAF, and CR. | |
| Q29 | Author: Please add page range in reference 3. | |
| Q30 | Author: Add page range in reference 15. | |
| Q31 | Author: Reference 28: Please clarify author's last name/first-name initials for "Treacher DF TR". | |
| Q32 | Author: References 35 and 43: Provide the first 10 authors followed by "et al" (if >10 authors). | |



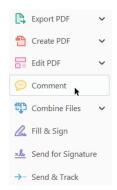
MAKING CORRECTIONS TO YOUR PROOF

These instructions show you how to mark changes or add notes to your proofs using Adobe Acrobat Professional versions 7 and onwards, or Adobe Reader DC. To check what version you are using go to **Help** then **About**. The latest version of Adobe Reader is available for free from get.adobe.com/reader.

DISPLAYING THE TOOLBARS

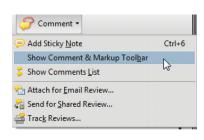
Adobe Reader DC

In Adobe Reader DC, the Comment toolbar can be found by clicking 'Comment' in the menu on the right-hand side of the page (shown below).



Acrobat Professional 7, 8, and 9

In Adobe Professional, the Comment toolbar can be found by clicking 'Comment(s)' in the top toolbar, and then clicking 'Show Comment & Markup Toolbar' (shown below).



The toolbar shown below will then be displayed along the top.

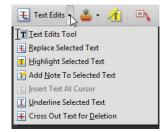


The toolbar shown below will then display along the top.



USING TEXT EDITS AND COMMENTS IN ACROBAT

This is the quickest, simplest and easiest method both to make corrections, and for your corrections to be transferred and checked.



- 1. Click **Text Edits**
- 2. Select the text to be annotated or place your cursor at the insertion point and start typing.
- 3. Click the **Text Edits** drop down arrow and select the required action.

You can also right click on selected text for a range of commenting options, or add sticky notes.

SAVING COMMENTS

In order to save your comments and notes, you need to save the file (File, Save) when you close the document.

USING COMMENTING TOOLS IN ADOBE READER

All commenting tools are displayed in the toolbar. You cannot use text edits, however you can still use highlighter, sticky notes, and a variety of insert/replace text options.

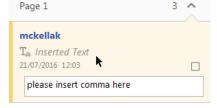


POP-UP NOTES

In both Reader and Acrobat, when you insert or edit text a pop-up box will appear. In **Acrobat** it looks like this:



In Reader it looks like this, and will appear in the right-hand pane:



DO NOT MAKE ANY EDITS DIRECTLY INTO THE TEXT, USE COMMENTING TOOLS ONLY.

Original Research Communications



Plasma trimethylamine-N-oxide and related metabolites are associated with type 2 diabetes risk in the Prevención con Dieta Mediterránea (PREDIMED) trial

Christopher Papandreou, ^{1,2} Mònica Bulló, ^{1,2} Yan Zheng, ³ Miguel Ruiz-Canela, ^{2,5} Edward Yu, ³ Marta Guasch-Ferré, ^{1,2,3} Estefanía Toledo, ^{2,5} Clary Clish, ⁶ Dolores Corella, ^{2,7} Ramon Estruch, ^{2,7,9} Emilio Ros, ^{2,10} Montserrat Fitó, ^{2,11} Fernando Arós, ^{2,12} Miquel Fiol, ^{2,13} José Lapetra, ^{2,14} Lluís Serra-Majem, ^{2,15} Enrique Gómez-Gracia, ¹⁶ Liming Liang, ⁴ Georgios A Fragkiadakis, ¹⁷ Cristina Razquin, ^{2,5} Frank B Hu, ^{3,4,18} and Jordi Salas-Salvadó ^{1,2}

Human Nutrition Unit, Faculty of Medicine and Health Sciences, Institut d'Investigació Sanitària Pere Virgili, Rovira i Virgili University, Reus, Spain;
 ²CIBER Fisiopatología de la Obesidad y Nutrición (CIBERObn), Instituto de Salud Carlos III, Madrid, Spain;
 ³Departments of Nutrition;
 ⁴Epidemiology and Statistics, Harvard TH Chan School of Public Health, Boston, MA;
 ⁵Department of Preventive Medicine and Public Health, University of Navarra, IdisNA, Pamplona, Spain;
 ⁶Broad Institute of MIT and Harvard University, Cambridge, MA;
 ⁷Department of Preventive Medicine, University of Valencia, Valencia, Spain;
 ⁸Departments of Internal Medicine;
 ⁹Endocrinology and Nutrition, Institut d'Investigacions Biomediques August Pi Sunyer (IDI-BAPS), Hospital Clinic, University of Barcelona, Barcelona, Spain;
 ¹⁰Lipid Clinic, Department of Endocrinology and Nutrition, IDI-BAPS, Hospital Clinic, University of Barcelona, Barcelona, Spain;
 ¹¹Cardiovascular and Nutrition Research Group, Institut de Recerca Hospital del Mar, Barcelona, Spain;
 ¹²Department of Cardiology, University Hospital of Alava, Vitoria, Spain;
 ¹³Institute of Health Sciences IUNICS, University of Balearic Islands and Hospital Son Espases, Palma de Mallorca, Spain;
 ¹⁴Department of Family Medicine, Primary Care Division of Sevilla, San Pablo Health Center, Seville, Spain;
 ¹⁵Department of Clinical Sciences, University of Las Palmas de Gran Canaria, Las Palmas, Spain;
 ¹⁶Department of Preventive Medicine, University of Málaga, Malaga, Spain;
 ¹⁷Department of Nutrition and Dietetics, Technological Education Institute of Crete, Siteia, Crete, Greece; and ¹⁸Channing Division for Network Medicine, Department of Medicine, Brigham and Women's Hospital and Harvard Medical School, Boston, MA

ABSTRACT

Background: The role of trimethylamine-N-oxide (TMAO) in type 2 diabetes (T2D) is currently partially understood and controversial. **Objective:** The aim was to investigate associations between TMAO and related metabolites with T2D risk in subjects at high risk of cardiovascular disease.

Design: This is a case-cohort design study within the Prevención con Dieta Mediterránea (PREDIMED) study, with 251 incident T2D cases and a random sample of 694 participants (641 noncases and 53 overlapping cases) without T2D at baseline (median follow-up: 3.8 y). We used liquid chromatography—tandem mass spectrometry to measure plasma TMAO, L-carnitine, betaine, lyso-phosphatidylcholine (LPC) and lyso-phosphatidylethanolamine (LPE) species, phosphocholine, α -glycerophosphocholine, and choline at baseline and after 1 y. We examined associations with the use of weighted Cox proportional hazard models, accounting for the weighted case-cohort design by the Barlow method.

Results: After adjustment for recognized T2D risk factors and multiple testing, individuals in the highest quartile of baseline TMAO and α -glycerophosphocholine had a lower risk of T2D [HR (95% CI): 0.52 (0.29, 0.89) and 0.46 (0.24, 0.89), respectively]. The HR (95% CI) comparing the extreme quartiles of betaine was 0.41 (0.23, 0.74). Similar trends were observed for C16:0 LPC, C18:1 LPC, C18:0 LPC, C20:4 LPC, C22:6 LPC, C18:1 LPC plasmalogen, and C16:0 LPE. After correcting for multiple comparisons, participants in the highest quartile of 1-y changes in C18:1 LPC plasmalogen concentrations had a lower T2D risk than the reference quartile.

Conclusion: Whether the associations between plasma TMAO and certain metabolite concentrations with T2D risk reflect its pathophysiology or represent an epiphenomenon need to be elucidated. This trial is registered at http://www.controlled-trials.com as IS-RCTN35739639. *Am J Clin Nutr* 2018;0:1–10.

Keywords: trimethylamine-N-oxide, metabolites, type 2 diabetes, case-cohort, Mediterranean diet, PREDIMED

INTRODUCTION

Plasma concentrations of trimethylamine-N-oxide (TMAO) are determined by diet, the gut microbiome, and liver flavin-containing monooxygenase 3 (FMO3) activities (1). They are directly associated with the consumption of animal-derived foods (2, 3) containing choline, phosphatidylcholine, and L-carnitine, which are processed by gut bacteria resulting in the release of various metabolites including trimethylamine (TMA), into the blood. TMA is then transported to the liver where it is converted to TMAO, which is involved in various physiologic and pathophysiologic processes such as the deposition and removal of cholesterol from the artery endothelium (4, 5).

Supported by research grant R01-DK-102896 from the NIH. The Prevención con Dieta Mediterránea (PREDIMED) trial was supported by the official funding agency for biomedical research of the Spanish government,

TMAO has been proposed as a molecule mediating the development of type 2 diabetes (T2D) (6). Higher plasma TMAO concentrations and alterations in interrelated pathways, such as phospholipid modification and methylation, have been associated with T2D (7). In this context, plasma choline concentrations have been found to be positively related, whereas betaine concentrations are inversely related to glucose concentrations (8) and lowered in insulin-resistant subjects (9). Betaine can be derived from choline or from L-carnitine metabolism (10), with L-carnitine associated with better insulin sensitivity in diabetics and with insulin-mediated glucose uptake in normoglycemic subjects (11).

Substantial inverse associations between dietary patterns consisting of healthy foods and the risk of T2D have been reported (12, 13). Recently, in a secondary outcome analysis of the Prevention of Disease with Mediterranean Diet [Prevención con Dieta Mediterránea (PREDIMED)] trial, a Mediterranean diet (MedDiet) reduced the risk of T2D by 30% compared with the control group (14). Although the benefits of this dietary pattern for T2D prevention is clearly observed in the PREDIMED trial, the biological mechanisms underlying these benefits are not completely understood. Consequently, an approach through metabolite-profiling technology (metabolomics) (15) can

the Instituto de Salud Carlos III, through grants provided to research networks specifically developed for the trial [grant RTIC G03/140 (to RE); grant RTIC RD 06/0045 (to MAM-G)] and through the Centro de Investigación Biomédica en Red de Fisiopatología de la Obesidad y Nutrición, and by grants from Centro Nacional de Investigaciones Cardiovasculares (grant CNIC 06/2007), the Fondo de Investigación Sanitaria Fondo Europeo de Desarrollo Regional (grants PI04-2239, PI 05/2584, CP06/00100, PI07/0240, PI07/1138, PI07/0954, PI 07/0473, PI10/01407, PI10/02658, PI11/01647, P11/02505, and PI13/00462), the Ministerio de Ciencia e Innovación (grants AGL-2009-13906-C02 and AGL2010-22319-C03), the Fundación Mapfre 2010, Consejería de Salud de la Junta de Andalucía (grant PI0105/2007), the Public Health Division of the Department of Health of the Autonomous Government of Catalonia, Generalitat Valenciana (grants ACOMP06109, GVA-COMP2010-181, GVACOMP2011-151, CS2010-AP-111, and CS2011-AP-042), and the Regional Government of Navarra (grant P27/2011). CP was supported by a postdoctoral fellowship granted by the Autonomous Government of Catalonia (PERIS 2016-2020; Incorporació de Científics I Tecnòlegs, SLT002/0016/00428). MG-F was supported by a postdoctoral fellowship granted by the Lilly Foundation European Association of Diabetes (EASD) through the Institut d'Investigacions Sanitàries Pere i Virgili (IISPV), Tarragona, Spain. The Fundación Patrimonio Comunal Olivarero and Hojiblanca (Malaga, Spain), the California Walnut Commission (Sacramento, California), Borges (Reus, Spain), and Morella Nuts (Reus, Spain) donated the olive oil, walnuts, almonds, and hazelnuts, respectively, used in the study; however, none of these funding sources played a role in the design, collection, analysis, interpretation or publication of the data.

Supplemental Figure 1 and Supplemental Tables 1–6 are available from the "Supplementary data" link in the online posting of the article and from the same link in the online table of contents at https://academic.oup.com/ajcn/.

Address correspondence to CP (e-mail: papchris10@gmail.com) or JS-S (e-mail: jordi.salas@urv.cat).

Abbreviations used: Ach, acetylcholine; CVD, cardiovascular disease; LPC, lyso-phosphatidylcholine; LPE, lyso-phosphatidylethanolamine; MedDiet, Mediterranean diet; PREDIMED, Prevención con Dieta Mediterránea; T2D, type 2 diabetes; TMA, trimethylamine; TMAO, trimethylamine-Novide

Received October 12, 2017. Accepted for publication March 7, 2018. First published online 0, 2018; doi: https://doi.org/10.1093/ajcn/nqy058.

strengthen existing pathophysiologic evidence, providing further support for dietary prevention of T2D.

To our knowledge, no prospective study has assessed the association between TMAO with T2D risk. Thus, the role of TMAO in T2D development is not completely understood. Taking this into account, the primary aim of the present prospective study, nested in the framework of the PREDIMED trial, was to examine possible associations between baseline and 1-y changes in the concentrations of TMAO and several metabolites involved in relevant pathways [L-carnitine, betaine, lyso-phosphatidylcholine (LPC) and lyso-phosphatidylethanolamine (LPE) species, phosphocholine, α -glycerophosphocholine, and choline], with the risk of incident T2D. In addition, we aimed to examine whether a MedDiet modified these associations.

METHODS

Study design and participants

This study used a case-cohort design nested within the PRED-IMED trial (ISRCTN35739639), a multicenter, single-blinded, controlled trial, conducted in Spanish primary health care centers. The design of the PREDIMED trial has been described in detail elsewhere (16, 17). In brief, 7447 participants at high cardiovascular disease (CVD) risk were allocated to a MedDiet supplemented with extra-virgin olive oil, a MedDiet supplemented with mixed nuts, or a control diet consisting of advice to reduce fat intake. For the present study, we considered the 3541 participants who were free of T2D at study inception. The present casecohort study comprises a random selection of 694 nondiabetic participants (?20%) from the eligible subjects of the PREDIMED cohort without T2D at study inception and with available blood samples, together with all incident cases of T2D that occurred during the follow-up with available plasma samples (251 out of the 273 cases). Of the 892 participants included in our analyses, 641 were in the subcohort (including 53 overlapping cases between the subcohort and the total cases) and 198 comprised the rest of the T2D cases, which gave a total of 251 cases (Supplemental Figure 1). Of these, 686 out of the 892 participants had available samples after 1 y of follow-up and were included in the 1-y change analyses (Supplemental Figure 1). The institutional review boards of the recruitment centers approved the study protocol, and participants provided written informed consent.

Study samples and metabolomics profiling

Fasting (for ≥ 8 h) plasma EDTA samples were collected from subjects and stored at -80° C. In June 2015, pairs of samples for each participant (baseline and at the end of the 1-y follow-up) were randomly ordered and shipped on dry ice to the Broad Institute, Inc., Boston, Massachusetts, for metabolomics assays. Liquid chromatography–tandem mass spectrometry techniques were used to perform semiquantitative profiling of several metabolites in blood plasma (TMAO, L-carnitine, betaine, LPC and LPE species, phosphocholine, α -glycerophosphocholine, and choline) and identify relations between them. A system composed of a Shimadzu Nexera X2 U-HPLC (Shimadzu Corp.) coupled to a Q Exactive hybrid quadrupole orbitrap mass spectrometer (Thermo Fisher Scientific) was used (18–24). Metabolite identities were confirmed by using authentic reference standards. Raw data were

Q٤

Q

processed with the use of TraceFinder software (Thermo Fisher Scientific) and Progenesis QI (Nonlinear Dynamics).

Ascertainment of T2D cases

Information was collected through contact with participants and primary health care physicians, annual follow-up visits, yearly ad hoc reviews of medical charts, and annual consultation of the National Death Index. Because T2D was a prespecified secondary outcome of the PREDIMED trial, it was identified at baseline by clinical diagnosis or use of antidiabetic medication. The diagnosis of new-onset T2D during follow-up has been described elsewhere (14) and briefly followed the American Diabetes Association criteria (25), namely 2 confirmations of fasting plasma glucose \geq 7.0 mmol/L or 2-h plasma glucose \geq 11.1 mmol/L after a 75-g oral-glucose load.

Assessment of covariates and other variables

At baseline and yearly during the follow-up, the participants completed a 47-item questionnaire related to lifestyle variables, smoking status, medical history, and medication use. A validated Spanish version of the Minnesota Leisure-Time Physical Activity Questionnaire was administered in order to evaluate physical activity (26). To assess the degree of adherence to the MedDiet, a 14-item validated questionnaire was filled in for each participant (27). BMI was calculated as weight divided by height squared (kg/m²). Participants' triacylglycerol and total, HDL, and LDL cholesterol concentrations were measured by using fasting plasma at baseline. Blood glucose and insulin concentrations were centrally assessed at baseline and at the end of the 1 y of follow-up. Insulin resistance was estimated by the HOMA-IR method with the use of the following equation (28): HOMA-IR = [fasting insulin (μ IU/mL) × fasting glucose (mmol/L)]/22.5.

Statistical analysis

0.10

Baseline characteristics of cases and noncases are described as means \pm SDs for quantitative variables and percentages or numbers for categorical variables. We applied a natural logarithmic transformation to approximate a normal distribution of metabolite concentrations. Person-time of follow-up was calculated as the interval between the baseline visit and date of T2D event, death, or date of the last contact, whichever came first. We used Cox proportional hazard models, with Barlow weights (to account for the overrepresentation of cases), to estimate HRs and their 95% CIs for the risk of T2D. A crude model and 2 multivariable-adjusted Cox regression models were fitted as follows: 1) multivariable model 1 adjusted for age (years), sex (male or female), BMI (kg/m²), intervention group, and baseline fasting glucose (milligrams per deciliter) (adding a quadratic term to account for the departure from linearity) and 2) multivariable model 2 additionally adjusted for smoking (never, current, or former), leisure-time physical activity (metabolic equivalent tasks in minutesper day), baseline dyslipidemia (yes or no), and hypertension (yes or no). We stratified the models according to recruitment center. Baseline metabolites were analyzed as both continuous variables (1-SD increment in their transformed levels) and by using quartiles (using cutoffs defined among noncases). To appraise

the linear trend across quartiles, the median metabolite concentration within each quartile was included in the Cox regression models as a continuous variable. To account for multiple testing, we adjusted P values of the multivariable-adjusted associations between quartiles or 1-SD increment in metabolite concentration and T2D risk with the use of the Benjamini-Hochberg false discovery rate procedure (29). A false discovery rate P value < 0.05 was considered to be significant.

We also examined the associations of 1-y changes in metabolites with T2D risk. We used the same models as in the baseline value analyses but further adjusted for baseline metabolite concentrations. With respect to metabolites, we first calculated the ratio between 1-y and baseline values and then normalized this ratio with the natural logarithmic transformation. To test the robustness of our results in relation to the association between TMAO and T2D risk, we conducted 2 sensitivity analyses: 1) testing the association between baseline values and T2D risk after excluding early cases (<1 y) and 2) testing the associations between the mean values at baseline and 1-y follow-up and subsequent T2D risk (T2D cases that occurred from baseline through the 1-y follow-up were excluded). To examine whether the association between baseline and 1-y changes in metabolites and incident T2D varied by intervention group, we stratified the analysis described above by intervention group (both merged MedDiet interventions compared with the control). We also added a multiplicative term (1 df) between intervention assignment (merged MedDiet compared with the control group) and metabolites (continuous) into the multivariable Cox models stratified on intervention assignment to test for interactions by means of likelihood ratio tests. In addition, we compared differences in 1-y changes in metabolites in the MedDiet group with changes in the control group (adjusted for the aforementioned covariates) with the use of ANCOVA. Finally, we applied multiple linear regression analyses to examine relations between TMAO concentrations and precursors (choline, betaine, and L-carnitine) at baseline and at the 1-y follow-up in the whole group, adjusting for age, sex, and HOMA-IR. Statistical analyses were performed with the use of Stata 13.1 (StataCorp.). A 2-sided P value < 0.05 was considered significant.

RESULTS

Baseline characteristics

The median follow-up of the study population was 3.8 y. The baseline characteristics of the 892 subjects (251 cases and 641 noncases) included in the present case-cohort study are shown in **Table 1**. The mean age of participants at baseline was 66.5 y, and the mean \pm SD BMI was 30.1 \pm 3.5. Compared with noncases, those participants who developed T2D were more likely to be men, current smokers, and to have a higher prevalence of hypertension in addition to a higher BMI and fasting glucose and triacylglycerol concentrations (Table 1).

Baseline metabolites and risk of T2D

Overall group

The associations between plasma metabolites with the risk of T2D in the overall group are presented in **Table 2**. In the group

12

TABLE 1Baseline characteristics of the study population¹

O16

| | Total | Cases | Noncases | P |
|---|-------------------|-------------------|-------------------|---------|
| \overline{n} | 892 | 251 | 641 | |
| Age, y | 66.5 ± 5.7 | 66.4 ± 5.7 | 66.5 ± 5.7 | 0.781 |
| Sex, % female | 61.2 | 55.0 | 63.6 | 0.017 |
| BMI, kg/m ² | 30.1 ± 3.5 | 30.8 ± 3.3 | 29.8 ± 3.6 | < 0.001 |
| Physical activity, METs/d | 240.7 ± 234.6 | 249.2 ± 233.5 | 237.4 ± 235.1 | 0.500 |
| Intervention group, % | | | | |
| MedDiet + EVOO | 30.6 | 29.9 | 30.9 | 0.425 |
| MedDiet + nuts | 36.3 | 37.2 | 33.8 | |
| Control group | 33.1 | 36.3 | 31.8 | |
| Hypertension, % | 91.7 | 96.0 | 90.0 | 0.003 |
| Dyslipidemia, % | 84.3 | 79.7 | 86.1 | 0.018 |
| Smoking, % | | | | |
| Never | 59.0 | 52.6 | 61.5 | 0.006 |
| Former | 22.4 | 22.3 | 22.5 | |
| Current | 18.6 | 25.1 | 16.0 | |
| Score for adherence to MedDiet ² | 8.5 ± 2.0 | 8.4 ± 2.0 | 8.6 ± 1.9 | 0.186 |
| Fasting blood glucose, mg/dL | 103.3 ± 17.6 | 118.6 ± 18.0 | 97.8 ± 13.8 | < 0.001 |
| Total cholesterol, mg/dL | 222.2 ± 39.3 | 221.7 ± 42.3 | 222.4 ± 38.1 | 0.846 |
| HDL cholesterol, mg/dL | 54.7 ± 13.0 | 52.6 ± 12.7 | 55.7 ± 13.1 | 0.014 |
| LDL cholesterol, mg/dL | 139.7 ± 32.9 | 137.0 ± 31.6 | 141.0 ± 33.3 | 0.201 |
| Triacylglycerol, mg/dL | 140.1 ± 85.0 | 169.0 ± 121.0 | 128.6 ± 62.0 | < 0.001 |

¹Values are means ± SDs unless otherwise indicated. Chi-square test was used for comparison of categorical variables and Student's *t* test was used for comparison of continuous variables. EVOO, extra-virgin olive oil; MedDiet, Mediterranean diet; MET, metabolic equivalent.

adjusted for age, sex, BMI, intervention group, glucose, smoking, leisure-time physical activity, dyslipidemia, and hypertension (multivariable model 2), the estimated HR for incident T2D reached significance only in the highest, compared with the lowest, quartile of TMAO concentrations (HR: 0.52; 95% CI: 0.29, 0.89). In both sensitivity analyses, TMAO concentrations in the highest quartile were significantly associated with lower T2D risk (a: HR: 0.52; 95% CI: 0.29, 0.93; b: HR: 0.44; 95% CI: 0.20, 0.96) compared with the lowest quartile. In the group adjusted for age, sex, BMI, intervention group, and glucose (multivariable model 1), the highest quartile of phosphocholine was associated with a lower risk of T2D (HR: 0.51; 95% CI: 0.27, 0.99), but these associations were no longer significant after further adjustment. With regard to betaine, the estimated HR for incident T2D in the highest compared with the lowest quartile was 0.41 (95% CI: 0.23, 0.74; P = 0.003). Concerning lyso-choline species, significant inverse associations with T2D incidence were observed for C16:0 LPC, C18:1 LPC, C18:0 LPC, C20:4 LPC, C22:6 LPC, and C18:1 LPC plasmalogen, either metabolites were modeled continuously (per 1 SD) or as quartiles (Table 2). Notably, per 1-SD increase in C18:1 LPC plasmalogen concentrations, a 54% lower risk (HR: 0.46; 95% CI: 0.35, 0.59) of T2D was observed (P < 0.001), and individuals in the highest quartile had an HR of 0.15 (95% CI: 0.06, 0.35; P < 0.001).

With regard to lyso-ethanolamine species, concentrations of C16:0 LPE (per 1-SD increase and highest quartile) were significantly associated with a reduced risk of T2D. Per 1-SD increase in L-carnitine, α -glycerophosphocholine, and betaine concentrations, a 19% (HR: 0.81; 95% CI: 0.67, 0.97; P = 0.025), 39% (HR: 0.61; 95% CI: 0.45, 0.83; P = 0.002), and 25% (HR: 0.75; 95% CI: 0.61, 0.91; P = 0.004) lower risk of T2D was found,

respectively. These associations remained significant after accounting for multiple comparisons.

MedDiet and control group

In stratified analyses by intervention group, the estimated HR for incident T2D in the highest compared with the lowest quartile of α -glycerophosphocholine concentrations was 0.27 (95% CI: 0.12, 0.62; P=0.002) in the MedDiet group (**Supplemental Table 1**) and 1.08 (95%: CI 0.14, 1.16; P>0.05) in the control group (**Supplemental Table 2**). In the MedDiet group, the HR associated with a 1-SD increment in α -glycerophosphocholine concentration was 0.55 (95% CI: 0.37, 0.82; P=0.003, P-interaction = 0.041) (**Figure 1**A, **Supplemental Table 3**), whereas in the control group no significant associations were observed (Figure 1B).

One-year changes in concentrations of metabolites and the risk of T2D

Associations between 1-y changes in metabolite concentrations across quartiles and the risk of T2D are shown in **Table 3**. In the highest quartile of increase in the concentrations of TMAO, a significant inverse association with T2D risk was found (HR: 0.49; 95% CI: 0.25, 0.98). Several lyso-choline species (C16.0 LPC, C18.1 LPC, and C18.0 LPC), including C18.1 LPC plasmalogen, C16.0 LPE, and α -glycerophosphocholine, were significantly associated with a decreased risk of T2D. After adjustment for multiple testing, only C18.1 LPC plasmalogen remained significant. We repeated the analyses with the use of a 1-SD increment in 1-y change in metabolite concentrations and,

(17

²This score is based on the 14-item dietary screener.

TABLE 2Associations of baseline individual metabolite eoneentrations with the risk of type 2 diabetes in the PREDIMED trial, 2003–2010, in the overall group.

1

| | Quartile of plasma metabolite concentration | | | | | P | | | | |
|-----------------------------|--|-------------------|-------------------|-------------------|---------|----------------------------------|-----------------------|------------|------------------|--|
| Metabolite | 1 | 2 | 3 | 4 | P-trend | FDR-Adjusted p value (Q4 vs. Q1) | HR per 1-SD increment | Unadjusted | FDR- adjusted | |
| Trimethylamine | N-oxic | le | | | | | | | | |
| Cases, n | 73 | 65 | 68 | 44 | | | | | | |
| Crude model | Ref | 0.98 (0.64, 1.53) | 1.01 (0.66, 1.56) | 0.60 (0.37, 0.98) | 0.039 | | 0.88 (0.75, 1.04) | 0.162 | | |
| MV1 | Ref | 0.96 (0.60, 1.54) | 0.95 (0.58, 1.56) | 0.51 (0.29, 0.88) | 0.012 | 0.022 | 0.86 (0.72, 1.04) | 0.123 | 0.070 | |
| MV2 | Ref | 0.98 (0.60, 1.59) | 0.91 (0.55, 1.51) | 0.52 (0.29, 0.89) | 0.012 | 0.032 | 0.83 (0.69, 1.01) | 0.059 | 0.078 | |
| Phosphocholine | 75 | 66 | 57 | 49 | | | | | | |
| Cases, <i>n</i> Crude model | Ref | 0.99 (0.64, 1.52) | 0.77 (0.49, 1.21) | 0.56 (0.32, 0.98) | 0.029 | | 0.90 (0.77, 1.06) | 0.224 | | |
| MV1 | Ref | 1.11 (0.68, 1.80) | 0.82 (0.48, 1.39) | 0.51 (0.27, 0.99) | 0.029 | | 0.88 (0.74, 1.05) | 0.224 | | |
| MV2 | Ref | 1.19 (0.71, 1.97) | 0.82 (0.48, 1.59) | 0.56 (0.29, 1.09) | 0.030 | 0.124 | 0.90 (0.75, 1.08) | 0.137 | 0.329 | |
| Choline | ICI | 1.17 (0.71, 1.77) | 0.07 (0.51, 1.50) | 0.30 (0.2), 1.0)) | 0.077 | 0.124 | 0.50 (0.75, 1.00) | 0.200 | 0.32) | |
| Cases, n | 52 | 67 | 63 | 65 | | | | | | |
| Crude model | Ref | 1.20 (0.75, 1.90) | 1.04 (0.63, 1.72) | 1.34 (0.83, 2.18) | 0.313 | | 1.03 (0.86, 1.23) | 0.729 | | |
| MV1 | Ref | 1.33 (0.80, 2.24) | 1.07 (0.61, 1.88) | 1.20 (0.70, 2.07) | 0.698 | | 0.94 (0.77, 1.14) | 0.538 | | |
| MV2 | Ref | 1.42 (0.84, 2.42) | 1.13 (0.63, 2.02) | 1.23 (0.71, 2.13) | 0.663 | 0.493 | 0.94 (0.77, 1.14) | 0.523 | 0.557 | |
| C14:0 LPC | 101 | 1.12 (0.01, 2.12) | 1.13 (0.03, 2.02) | 1.23 (0.71, 2.13) | 0.005 | 0.175 | 0.51 (0.77, 1.11) | 0.323 | 0.557 | |
| Cases, n | 72 | 65 | 55 | 56 | | | | | | |
| Crude model | Ref | 1.05 (0.68, 1.64) | 0.91 (0.57, 1.45) | 0.91 (0.57, 1.44) | 0.590 | | 1.01 (0.85, 1.19) | 0.910 | | |
| MV1 | Ref | 1.02 (0.61, 1.74) | 0.89 (0.53, 1.49) | 0.93 (0.55, 1.55) | 0.669 | | 1.03 (0.86, 1.24) | 0.741 | | |
| MV2 | Ref | 1.05 (0.62, 1.78) | 0.88 (0.52, 1.49) | 0.88 (0.52, 1.50) | 0.548 | 0.650 | 1.01 (0.84, 1.23) | 0.879 | 0.879 | |
| C16:1 LPC | | , , , | | ` ' ' | | | , , , | | | |
| Cases, n | 78 | 61 | 58 | 50 | | | | | | |
| Crude model | Ref | 0.89 (0.57, 1.38) | 0.84 (0.54, 1.31) | 0.72 (0.45, 1.16) | 0.181 | | 0.91 (0.77, 1.08) | 0.317 | | |
| MV1 | Ref | 0.77 (0.47, 1.26) | 0.93 (0.57, 1.52) | 0.75 (0.45, 1.26) | 0.387 | | 0.91 (0.76, 1.11) | 0.367 | | |
| MV2 | Ref | 0.77 (0.47, 1.28) | 1.03 (0.63, 1.68) | 0.75 (0.44, 1.27) | 0.451 | 0.318 | 0.93 (0.77, 1.12) | 0.461 | 0.526 | |
| C16:0 LPC | | | | | | | | | | |
| Cases, n | 102 | 62 | 51 | 36 | | | | | | |
| Crude model | Ref | 0.60 (0.39, 0.92) | 0.58 (0.37, 0.91) | 0.41 (0.24, 0.69) | 0.001 | | 0.69 (0.56, 0.83) | < 0.001 | | |
| MV1 | Ref | 0.53 (0.33, 0.85) | 0.58 (0.36, 0.93) | 0.41 (0.23, 0.74) | 0.002 | | 0.69 (0.56, 0.84) | < 0.001 | | |
| MV2 | Ref | 0.50 (0.31, 0.82) | 0.56 (0.35, 0.92) | 0.42 (0.24, 0.75) | 0.002 | 0.012 | 0.69 (0.56, 0.85) | < 0.001 | 0.003 | |
| C18:1 LPC | | | | | | | | | | |
| Cases, n | 111 | 59 | 48 | 30 | | | | | | |
| Crude model | Ref | 0.52 (0.34, 0.79) | 0.44 (0.28, 0.68) | 0.31 (0.18, 0.51) | < 0.001 | | 0.66 (0.56, 0.77) | < 0.001 | | |
| MV1 | Ref | 0.51 (0.32, 0.82) | 0.50 (0.32, 0.80) | 0.34 (0.19, 0.60) | < 0.001 | 0.000 | 0.67 (0.56, 0.80) | < 0.001 | 0.002 | |
| MV2 | Ref | 0.52 (0.32, 0.85) | 0.54 (0.33, 0.86) | 0.36 (0.20, 0.65) | < 0.001 | 0.008 | 0.69 (0.58, 0.82) | < 0.001 | 0.003 | |
| C18:0 LPC | 100 | 64 | 40 | 20 | | | | | | |
| Cases, n | 108 | 64 | 48 | 30 | 0.001 | | 0.50 (0.40, 0.74) | 0.001 | | |
| Crude model | Ref | 0.55 (0.36, 0.84) | 0.42 (0.27, 0.66) | 0.37 (0.21, 0.64) | < 0.001 | | 0.59 (0.48, 0.74) | < 0.001 | | |
| MV1 | Ref | 0.59 (0.37, 0.96) | 0.48 (0.29, 0.79) | 0.34 (0.18, 0.68) | < 0.001 | 0.012 | 0.63 (0.50, 0.79) | < 0.001 | 0.002 | |
| MV2 C20:4 LPC | Ref | 0.58 (0.36, 0.95) | 0.47 (0.28, 0.79) | 0.37 (0.19, 0.74) | 0.001 | 0.013 | 0.65 (0.51, 0.82) | < 0.001 | 0.003 | |
| Cases, n | 103 | 56 | 47 | 43 | | | | | | |
| Crude model | Ref | 0.52 (0.34, 0.80) | 0.48 (0.31, 0.76) | 0.45 (0.28, 0.73) | < 0.001 | | 0.67 (0.56, 0.80) | < 0.001 | | |
| MV1 | Ref | 0.49 (0.30, 0.78) | 0.54 (0.33, 0.87) | 0.43 (0.25, 0.75) | 0.001 | | 0.66 (0.54, 0.80) | < 0.001 | | |
| MV2 | Ref | 0.48 (0.29, 0.79) | 0.57 (0.35, 0.93) | 0.44 (0.25, 0.78) | 0.003 | 0.013 | 0.67 (0.54, 0.81) | < 0.001 | 0.003 | |
| C22:6 LPC | 1101 | 0.10 (0.2), 0.7) | 0.07 (0.00, 0.50) | 0111 (0120, 0170) | 0.002 | 0.015 | 0.07 (0.0 1, 0.01) | 10.001 | 0.002 | |
| Cases, n | 93 | 72 | 52 | 33 | | | | | | |
| Crude model | Ref | 1.03 (0.68, 1.55) | 0.71 (0.47, 1.09) | 0.39 (0.23, 0.66) | < 0.001 | | 0.76 (0.65, 0.89) | 0.001 | | |
| MV1 | Ref | 1.12 (0.70, 1.79) | 0.98 (0.63, 1.54) | 0.40 (0.22, 0.74) | 0.009 | | 0.78 (0.66, 0.94) | 0.007 | | |
| MV2 | Ref | 1.13 (0.71, 1.82) | 1.07 (0.67, 1.70) | 0.41 (0.22, 0.77) | 0.022 | 0.014 | 0.80 (0.67, 0.96) | 0.016 | 0.028 | |
| C18:1 LPC plasr | | | (, | (** , ****, | | | (,, | | | |
| Cases, n | 120 | 61 | 48 | 14 | | | | | | |
| Crude model | Ref | 0.56 (0.37, 0.86) | 0.38 (0.24, 0.59) | 0.18 (0.09, 0.37) | < 0.001 | | 0.42 (0.33, 0.55) | < 0.001 | | |
| MV1 | Ref | 0.68 (0.43, 1.09) | 0.44 (0.27, 0.72) | 0.14 (0.06, 0.32) | < 0.001 | | 0.45 (0.34, 0.58) | < 0.001 | | |
| MV2 | Ref | 0.67 (0.42, 1.08) | 0.45 (0.27, 0.74) | 0.15 (0.06, 0.35) | < 0.001 | 0.008 | 0.46 (0.35, 0.59) | < 0.001 | 0.003 | |
| C16:0 LPE | | | | | | | | | | |
| Cases, n | 94 | 73 | 50 | 32 | | | | | | |
| Crude model | Ref | 0.97 (0.64, 1.45) | 0.59 (0.38, 0.92) | 0.45 (0.26, 0.78) | 0.001 | | 0.73 (0.61, 0.87) | 0.001 | | |
| MV1 | Ref | 0.86 (0.54, 1.36) | 0.56 (0.35, 0.89) | 0.49 (0.27, 0.88) | 0.003 | | 0.74 (0.61, 0.89) | 0.002 | | |
| MV2 | Ref | 0.89 (0.55, 1.44) | 0.55 (0.34, 0.88) | 0.46 (0.25, 0.84) | 0.002 | 0.026 | 0.73 (0.60, 0.89) | 0.002 | 0.004 | |

TABLE 2
Continued.

| | Quartile of plasma metabolite concentration | | | | P | | | | |
|-----------------|---|-------------------|-------------------|-------------------|---------|--|-----------------------|------------|------------------|
| Metabolite | 1 | 2 | 3 | 4 | P-trend | nd FDR-Adjusted p value (Q4 vs. Q1) | HR per 1-SD increment | Unadjusted | FDR- adjusted |
| C18:1 LPE | | | | | | | | | |
| Cases, n | 73 | 66 | 61 | 49 | | | | | |
| Crude model | Ref | 0.89 (0.57, 1.38) | 0.84 (0.54, 1.30) | 0.60 (0.37, 0.95) | 0.034 | | 0.81 (0.69, 0.95) | 0.011 | |
| MV1 | Ref | 0.77 (0.47, 1.26) | 0.79 (0.50, 1.26) | 0.57 (0.34, 0.96) | 0.045 | | 0.82 (0.70, 0.99) | 0.040 | |
| MV2 | Ref | 0.79 (0.48, 1.32) | 0.86 (0.53, 1.39) | 0.59 (0.35, 1.00) | 0.073 | 0.075 | 0.84 (0.69, 1.00) | 0.058 | 0.078 |
| L-Carnitine | | | | | | | | | |
| Cases, n | 52 | 79 | 69 | 49 | | | | | |
| Crude model | Ref | 1.81 (1.12, 2.91) | 1.44 (0.89, 2.34) | 0.91 (0.54, 1.55) | 0.379 | | 0.90 (0.77, 1.05) | 0.198 | |
| MV1 | Ref | 1.76 (1.07, 2.90) | 1.09 (0.64, 1.88) | 0.75 (0.41, 1.36) | 0.108 | | 0.84 (0.70, 1.01) | 0.071 | |
| MV2 | Ref | 1.77 (1.06, 2.94) | 1.07 (0.61, 1.88) | 0.69 (0.37, 1.26) | 0.066 | 0.280 | 0.81 (0.67, 0.97) | 0.025 | 0.040 |
| α-Glycerophospl | hochol | ine | | | | | | | |
| Cases, n | 81 | 56 | 71 | 40 | | | | | |
| Crude model | Ref | 0.70 (0.44, 1.12) | 0.81 (0.51, 1.28) | 0.57 (0.32, 1.02) | 0.091 | | 0.70 (0.54, 0.92) | 0.011 | |
| MV1 | Ref | 0.58 (0.35, 0.98) | 0.75 (0.46, 1.20) | 0.47 (0.25, 0.90) | 0.044 | | 0.61 (0.45, 0.81) | 0.001 | |
| MV2 | Ref | 0.54 (0.31, 0.93) | 0.75 (0.46, 1.23) | 0.46 (0.24, 0.89) | 0.051 | 0.035 | 0.61 (0.45, 0.83) | 0.002 | 0.004 |
| Betaine | | | | | | | | | |
| Cases, n | 76 | 69 | 59 | 43 | | | | | |
| Crude model | Ref | 0.82 (0.53, 1.26) | 0.82 (0.52, 1.29) | 0.48 (0.29, 0.78) | 0.005 | | 0.83 (0.69, 0.98) | 0.035 | |
| MV1 | Ref | 0.90 (0.57, 1.43) | 0.74 (0.45, 1.21) | 0.41 (0.23, 0.73) | 0.002 | | 0.77 (0.64, 0.94) | 0.009 | |
| MV2 | Ref | 0.87 (0.54, 1.39) | 0.73 (0.43, 1.24) | 0.41 (0.23, 0.74) | 0.003 | 0.012 | 0.75 (0.61, 0.91) | 0.004 | 0.008 |

¹Values are HRs (95% CIs) unless otherwise indicated. A natural logarithmic transformation was applied to the raw values of individual metabolites. Cox regression analysis was used. MV1 adjusted for age (years), sex (male or female), BMI (kg/m²), intervention group (MedDiet + EVOO or MedDiet + nuts), and baseline fasting glucose (milligrams per deciliter) (centered on the sample mean and adding the quadratic term); MV2 additionally adjusted for smoking (never, current, or former), leisure-time physical activity (metabolic equivalent tasks in minutes per day), dyslipidemia, and hypertension. FDR-controlled adjustments were conducted by applying the method of Benjamini and Hochberg. EVOO, extra-virgin olive oil; FDR, false discovery rate; LPC, lyso-phosphatidylcholine; LPE, lyso-phosphatidylethanolamine; MedDiet, Mediterranean diet; MV, multivariable model; PREDIMED, Prevención con Dieta Mediterránea; Q, quartile; Ref, reference.

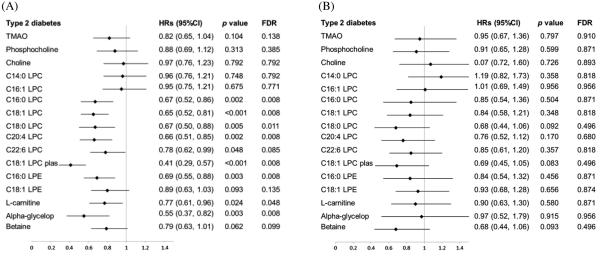


FIGURE 1 HRs (95% CIs) for type 2 diabetes according to baseline metabolites analyzed as continuous (per 1-SD increment) by both Mediterranean diet intervention groups (merged) (A) and the control group (B). All HRs were adjusted for age (y), sex (male or female), BMI (kg/m²), baseline fasting glucose (mg/dL; mean + quadratic term of the centered mean), smoking (never, current, or former), leisure-time physical activity (metabolic equivalent tasks in min/d), dyslipidemia, and hypertension. A natural logarithmic transformation was applied to the raw value. FDR-controlled adjustments were conducted by applying the method of Benjamini and Hochberg. Alpha-glycelop, α-glycerophosphocholine; FDR, false discovery rate; LPC, lysophosphatidylcholine; LPE, lysophosphatidylethanolamine; plas, plasmalogen; TMAO, trimethylamine N-oxide.

after adjustment for multiple testing, we found that per 1-SD increase in C16:0 LPC, C18:1 LPC, C18:0 LPC, and C18:1 LPC plasmalogen concentrations, the risk of T2D decreased by 35% (HR: 0.65; 95% CI: 0.49, 0.86), 37% (HR: 0.63; 95% CI: 0.47,

0.85), 32% (HR: 0.68; 95% CI: 0.50, 0.92), and 42% (HR: 0.58; 95% CI: 0.42, 0.81), respectively. These significant inverse associations observed in the overall group persisted in the MedDiet group, whereas only C18:1 LPC was found to be significantly

TABLE 3Associations of 1-y changes in individual metabolite concentrations with the risk of type 2 diabetes in the PREDIMED trial, 2003–2010, in the overall group 1

| | | Quartile of plas | | | | |
|-------------------------|----------|-------------------|-------------------|--------------------|---------|----------------------------|
| Metabolite | 1 | 2 | 3 | 4 | P-trend | FDR-adjusted P (Q4 vs. Q1) |
| Trimethylamine N | N-oxide | | | | | |
| Cases, n | 44 | 44 | 45 | 48 | | |
| MV | Ref | 0.58 (0.31, 1.10) | 0.64 (0.32, 1.25) | 0.49 (0.25, 0.98) | 0.081 | 0.100 |
| Phosphocholine | | | | | | |
| Cases, n | 44 | 44 | 45 | 48 | | |
| MV | Ref | 0.75 (0.42, 1.36) | 1.00 (0.59, 1.71) | 0.74 (0.40, 1.37) | 0.484 | 0.598 |
| Choline | | | | | | |
| Cases, n | 49 | 44 | 50 | 37 | | |
| MV | Ref | 1.11 (0.61, 2.02) | 0.83 (0.46, 1.51) | 1.04 (0.54, 2.00) | 0.895 | 0.924 |
| C14:0 LPC | | | | | | |
| Cases, n | 52 | 47 | 39 | 43 | | |
| MV | Ref | 0.77 (0.42, 1.40) | 0.79 (0.43, 1.47) | 0.79 (0.38, 1.64) | 0.587 | 0.769 |
| C16:1 LPC | | , , , | ` ' ' | ` ' ' | | |
| Cases, n | 57 | 35 | 43 | 45 | | |
| MV | Ref | 0.47 (0.24, 0.91) | 0.60 (0.33, 1.10) | 0.86 (0.44, 1.68) | 0.657 | 0.811 |
| C16:0 LPC | 1101 | 0 (0.2., 0.5.1) | 0.00 (0.00, 1.10) | 0.00 (0.1.1, 1.00) | 0.027 | 0.011 |
| Cases, n | 55 | 44 | 42 | 40 | | |
| MV | Ref | 0.57 (0.31, 1.04) | 0.38 (0.21, 1.69) | 0.47 (0.24, 0.93) | 0.014 | 0.100 |
| C18:1 LPC | ICI | 0.57 (0.51, 1.04) | 0.50 (0.21, 1.07) | 0.47 (0.24, 0.75) | 0.014 | 0.100 |
| Cases, n | 44 | 53 | 36 | 44 | | |
| MV | Ref | 0.76 (0.42, 1.38) | 0.55 (0.29, 1.01) | 0.44 (0.21, 0.90) | 0.016 | 0.100 |
| C18:0 LPC | ICI | 0.70 (0.42, 1.30) | 0.55 (0.25, 1.01) | 0.44 (0.21, 0.70) | 0.010 | 0.100 |
| Cases, n | 49 | 45 | 51 | 34 | | |
| MV | Ref | 0.63 (0.36, 1.12) | 0.47 (0.25, 0.90) | 0.45 (0.22, 0.92) | 0.018 | 0.100 |
| C20:4 LPC | Kei | 0.03 (0.30, 1.12) | 0.47 (0.23, 0.90) | 0.43 (0.22, 0.92) | 0.016 | 0.100 |
| | 48 | 41 | 42 | 49 | | |
| Cases, n MV | | | 0.53 (0.27, 1.03) | | 0.047 | 0.130 |
| | Ref | 0.74 (0.41, 1.35) | 0.33 (0.27, 1.03) | 0.54 (0.28, 1.04) | 0.047 | 0.130 |
| C22:6 LPC | 20 | 4.1 | 50 | 50 | | |
| Cases, n | 38 | 41 | 50 | 50 | 0.224 | 0.500 |
| MV | Ref | 0.99 (0.55, 1.79) | 0.55 (0.27, 1.10) | 0.72 (0.36, 1.47) | 0.334 | 0.598 |
| C18:1 LPC plasm | - | | 40 | 22 | | |
| Cases, n | 52 | 57 | 40 | 32 | 0.004 | 0.000 |
| MV | Ref | 0.76 (0.43, 1.33) | 0.30 (0.15, 0.58) | 0.33 (0.17, 0.66) | < 0.001 | 0.032 |
| C16:0 LPE | | | | | | |
| Cases, n | 44 | 51 | 45 | 34 | | |
| MV | Ref | 0.87 (0.48, 1.58) | 0.91 (0.51, 1.62) | 0.49 (0.25, 0.98) | 0.052 | 0.100 |
| C18:1 LPE | | | | | | |
| Cases, n | 42 | 50 | 32 | 56 | | |
| MV | Ref | 1.19 (0.66, 2.12) | 0.69 (0.38, 1.27) | 0.85 (0.43, 1.67) | 0.408 | 0.811 |
| L-Carnitine | | | | | | |
| Cases, n | 35 | 42 | 61 | 42 | | |
| MV | Ref | 1.16 (0.58, 2.32) | 2.05 (1.03, 4.11) | 0.86 (0.38, 1.91) | 0.725 | 0.811 |
| α -Glycerophosph | ocholine | | | | | |
| Cases, n | 49 | 47 | 51 | 33 | | |
| MV | Ref | 0.40 (0.21, 0.78) | 0.43 (0.22, 0.83) | 0.45 (0.21, 0.98) | 0.084 | 0.100 |
| Betaine | | | | | | |
| Cases, n | 41 | 47 | 45 | 48 | | |
| MV | Ref | 1.41 (0.77, 2.59) | 0.90 (0.47, 1.73) | 1.03 (0.54, 1.95) | 0.732 | 0.924 |

¹Values are HRs (95% CIs). We first calculated the ratio between 1-y and baseline concentrations of individual metabolites and then normalized this ratio with the natural logarithmic transformation. Cox regression analysis was used. The MV was adjusted for baseline concentrations of metabolites, age (years), sex (male or female), intervention group (MedDiet + EVOO or MedDiet + nuts), BMI (kg/m²), baseline fasting glucose (milligrams per deciliter) (centered on the sample mean and adding the quadratic term), smoking (never, current, or former), leisure-time physical activity (metabolic equivalent tasks in minutes per day), dyslipidemia, and hypertension. FDR-controlled adjustments were conducted by applying the method of Benjamini and Hochberg. EVOO, extra-virgin olive oil; FDR, false discovery rate; LPC, lyso-phosphatidylcholine; LPE, lyso-phosphatidylethanolamine; MedDiet, Mediterranean diet; MV, multivariable model; PREDIMED, Prevención con Dieta Mediterránea; Q, quartile; Ref, reference.

Q19

Q20

Q21

022

 Ω_{23}

Ω24

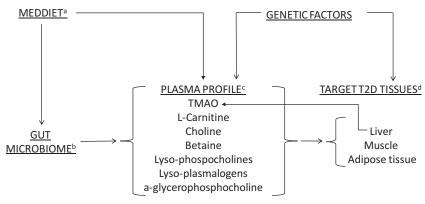


FIGURE 2 Open homeostasis pattern. ^aThe role of the Mediterranean diet; ^bthe role of the microbiome; ^c metabolites absorbed, produced and transformed, or methylated; ^dthe role of T2D pathogenicity target tissues. MEDDIET, Mediterranean diet; T2D, type 2 diabetes; TMAO, trimethylamine-N-oxide.

associated with lower risk in the control group. However, *P*-interaction values between C16:0 LPC, C18:1 LPC, C18:0 LPC, and the intervention group (MedDiet compared with the control group) and T2D were nonsignificant (**Supplemental Table 4**). There was a tendency toward higher T2D risk for those individuals in the control group with higher 1-y changes in betaine concentrations, but the results were not significant (HR: 1.25; 95% CI: 0.81, 1.92; *P*-interaction = 0.012). There were no significant differences in mean 1-y changes in metabolite concentrations between the MedDiet and control groups (data not shown).

Predictors of TMAO concentrations from regression analyses

A multiple linear regression was used to assess potential relations between TMAO and choline, betaine, and L-carnitine, while adjusting for age, sex, and HOMA-IR (**Supplemental Tables 5** and **6**). Baseline and 1-y changes in L-carnitine concentrations were positively associated with TMAO concentrations at baseline and at 1 y, respectively.

DISCUSSION

With the use of a case-cohort design within the PREDIMED trial and aimed at identifying plasma metabolites potentially related to T2D in 892 individuals at high CVD risk, we observed that higher baseline concentrations of TMAO, L-carnitine, betaine, α -glycerophosphocholine, and several LPC and LPE species were associated with lower risk of T2D development, independently of recognized T2D risk factors (i.e., age, sex, BMI, blood glucose, smoking). At baseline, the participants in the highest quartile of TMAO plasma concentrations had a 48% lower risk of developing T2D compared with the lowest quartile. Notably, the association between TMAO and T2D persisted and remained significant after sensitivity analyses.

L-Carnitine affects insulin-mediated glucose uptake and oxidation in diabetics and healthy controls, improving insulin sensitivity and blood glucose concentrations in patients with T2D (11) and mitochondrial utilization of fatty acids (30). Betaine is associated with a lower risk of diabetes, presumably because it serves as a methyl donor in the methionine cycle and its administration decreases homocysteine concentrations (31–33). Recently, plasma betaine concentrations were reported to be reduced in

insulin-resistant humans and were associated with insulin sensitivity (9). In addition, lower plasma concentrations of betaine were reported in patients with metabolic syndrome compared with a healthy population (34). The possible protective role of L-carnitine and betaine in T2D development was also clearly supported by our findings.

The gut microbiome is involved in gastrointestinal and immune function as well as the digestion of nutrients, and may thus affect the risk of obesity and T2D (35). L-Carnitine, choline, and betaine can be metabolized by the microbiome to TMA, which is absorbed by the gut and further oxidized to TMAO in the liver (Figure 2). Among these substrates, L-carnitine was identified in our study as the sole positive predictor of plasma TMAO coneentrations (10, 36). L-Carnitine may be transformed to TMAO through ≥2 independent pathways catalyzed by L-carnitine dehydrogenase or carnitine oxygenase or reductase (10). TMAO is, on the other hand, considered to be a risk factor for CVD (36). However, circulating TMAO is influenced by the gut microbiome, kidney function, as well as a FMO3 genotype, factors that may confound the relation between TMAO and chronic disease (37). Whether TMAO is a causal factor for disease development and progression or simply a biomarker of metabolic adaptation is unclear and needs to be explored further.

With regard to the inverse associations between TMAO plasma concentrations and the risk of developing T2D, our findings are compatible with a recent prospective study (38) involving a consecutive sample of 37 obese subjects undergoing bariatric surgery, 17 of whom had diabetes. One year after surgery, TMAO plasma concentrations increased by $\frac{9}{2}$ -fold compared with preoperative concentrations, which was significantly correlated with the corresponding 1-y decrease in glycated hemoglobin after the bariatric surgery (r = -0.39, P = 0.025). In our study, Spearman's correlation analysis between TMAO and 1-y increase in HOMA-IR, a value used to quantify insulin resistance, showed a significant negative correlation (r = -0.13, P = 0.026).

On the other hand, a study conducted in 283 individuals that examined cardiometabolic risk factors and pathways associated with TMAO concentration reported that the presence of diabetes was associated with higher concentrations of plasma TMAO (7). Similarly, Shan et al. (39) measured plasma concentrations of TMAO in a case-control study in 2694 participants and noted that higher plasma TMAO was associated with increased odds

25

of T2D. In addition, in a larger study in 4007 patients undergoing coronary angiography, increased plasma concentrations of TMAO were related to increased concentrations of fasting glucose and diabetes (40). These studies (7, 39, 40) were not designed to investigate associations between TMAO and T2D prospectively, whereas our data showed that higher TMAO concentrations can be prospectively associated with decreased T2D risk. However, differences in study design and the populations studied can only partly explain the overall discrepancies. Because TMAO was reported to be highly variable in plasma and urine (log-normal reference change values ranging from 403% to 80% in plasma) of overweight people with T2D over a 2-y period (41), further research is needed to explain these variations. Overall, the exact mechanisms associating plasma TMAO concentrations and T2D remain unclear (10). On the other hand, the possibility that changes in TMAO just represent an epiphenomenon cannot be excluded.

TMAO and choline concentration changes were linked to risk profiles with altered concentrations of phospholipids and methylation markers, with total or specific LPC concentrations tending to be lower among diabetics (7, 42, 43). LPC may play an important role in glucose homeostasis by stimulating adipocyte glucose uptake, potentiating glucose-stimulated insulin secretion and lowering blood glucose concentrations (43, 44). In our study, Spearman's correlations between baseline metabolite concentrations and 1-y changes in HOMA-IR values indicated a significant negative correlation between C18:1 LPC plasmalogen and HOMA-IR (r = -0.14, P = 0.010). Choline plasmalogens with 18:1 in the sn-2 position are strongly correlated with a wide range of risk factors for metabolic syndrome (45). Nonalcoholic fatty liver disease shares common characteristics with metabolic syndrome, such as insulin resistance (46), and has been associated with decreased plasma plasmalogens (47). In our study, higher C18:1 LPC plasmalogen plasma concentrations may reflect a decrease in insulin resistance during follow-up and a potential improvement in liver function, resulting in increased secretion of plasmalogens into the circulation. Because the role of LPCs in diabetes pathophysiology is not completed understood, we cannot exclude the possibility that they do not offer any protection against the disease.

Another choline compound, α -glycerophosphocholine, which in our study was found to be associated with a lower risk of T2D, can produce phosphorylcholine and acetylcholine (Ach), a neurotransmitter responsible for initiating skeletal muscle contractions (48). Activation of muscarinic Ach receptors in skeletal muscle cells stimulates glucose uptake via a mechanism independent of the insulin-stimulated pathway (49). Because all muscle movements are related to contraction, and contraction is related to available Ach stores, maximizing Ach may optimize muscular performance and affect insulin sensitivity, GLUT4 availability for glucose uptake, etc.

We did not observe any significant difference in the association of most 1-y change metabolites with T2D risk between the MedDiet intervention and control groups. Furthermore, the intervention diets did not appear to significantly change the study metabolite concentrations during the intervention. We found a stronger inverse association only for α -glycerophosphocholine with T2D risk in the MedDiet intervention compared with the control group with a significant test of this interaction. This finding needs further investigation.

The results of the present study should be interpreted in the context of its limitations and strengths. First, participants were elderly Mediterranean individuals at high CVD risk and this may limit the generalizability of the findings to other age groups or populations. Second, even though we adjusted for several potential confounders, residual confounding may exist. With regard to strengths, the prospective evaluation of the association between metabolite concentrations and incident T2D, in the frame of a case-cohort design, minimizes biases that can affect case-control studies.

In conclusion, our study documented, for the first time to our knowledge, a strong inverse prospective association between plasma TMAO concentrations and relevant metabolite profile with incident T2D risk in an elderly population at high CVD risk. Because TMAO is a metabolite that has shown controversial associations with several chronic diseases, these results may be reported in the literature to avoid bias but must be interpreted cautiously and need to be replicated in the future in other populations. The potential mechanisms linking the aforementioned metabolites and diabetes risk must also be further investigated.

The authors' responsibilities were as follows—FBH and JS-S: designed the research; CP, MB, MR-C, MG-F, ET, DC, RE, ER, M Fitó, FA, M Fiol, JL, LS-M, EG-G, and JS-S: conducted the research; DC, RE, M Fitó, FA, M Fiol, JL, LS-M, EG-G, and JS-S: were the coordinators of subject recruitment at the outpatient clinics; CP and MB: analyzed the data; CP, MB, FBH, and JS-S: performed the statistical analysis and interpreted the data; CC: analyzed metabolomics data; CP: drafted the manuscript; FBH and JS-S: supervised the study; CP, MB, and JS-S: had full access to all of the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis; and all authors are ded the manuscript. The authors declared that they had no conflicts of interest related to this article.

REFERENCES

- Velasquez MT, Ramezani A, Manal A, Raj DS. Trimethylamine Noxide: the good, the bad and the unknown. Toxins (Basel) 2016:8:326.
- Rohrmann S, Linseisen J, Allenspach M, von Eckardstein A, Müller D. Plasma concentrations of trimethylamine-N-oxide are directly associated with dairy food consumption and low-grade inflammation in a German adult population. J Nutr 2016;146:283–9.
- Cho CE, Taesuwan S, Malysheva OV, Bender E, Tulchinsky NF, Yan J, Sutter JL, Caudill MA. Trimethylamine-N-oxide (TMAO) response to animal source foods varies among healthy young men and is influenced by their gut microbiota composition: a randomized controlled trial. Mol Nutr Food Res 2017:61.
- Wang Z, Klipfell E, Bennett BJ, Koeth R, Levison BS, Dugar B, Feldstein AE, Britt EB, Fu X, Chung YM, et al. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. Nature 2011;472:57–63.
- Bennett BJ, de Aguiar Vallim TQ, Wang Z, Shih DM, Meng Y, Gregory J, Allayee H, Lee R, Graham M, Crooke R, et al. Trimethylamine-Noxide, a metabolite associated with atherosclerosis, exhibits complex genetic and dietary regulation. Cell Metab 2013;17:49–60.
- Kim Y, Keogh J, Clifton P. A review of potential metabolic etiologies of the observed association between red meat consumption and development of type 2 diabetes mellitus. Metabolism 2015;64:768–79.
- Obeid R, Awwad HM, Rabagny Y, Graeber S, Herrmann W, Geisel J. Plasma trimethylamine N-oxide concentration is associated with choline, phospholipids, and methyl metabolism. Am J Clin Nutr 2016;103:703–11.
- Konstantinova SV, Tell GS, Vollset SE, Nygård O, Bleie Ø, Ueland PM.
 Divergent associations of plasma choline and betaine with components
 of metabolic syndrome in middle age and elderly men and women. J
 Nutr 2008:138:914–20.
- Ejaz A, Martinez-Guino L, Goldfine AB, Ribas-Aulinas F, De Nigris V, Ribó S, Gonzalez-Franquesa A, Garcia-Roves PM, Li E, Dreyfuss

28

Q30

Q31

- JM, et al. Dietary betaine supplementation increases Fgf21 levels to improve glucose homeostasis and reduce hepatic lipid accumulation in mice. Diabetes 2016;65:902–12.
- Fennema D, Phillips IR, Shephard EA. Trimethylamine and trimethylamine N-oxide, a flavin-containing monooxygenase 3 (FMO3)-mediated host-microbiome metabolic axis implicated in health and disease. Drug Metab Dispos 2016;44:1839–50.
- Mingrone G, Greco AV, Capristo E, Benedetti G, Giancaterini A, De Gaetano A, Gasbarrini G. L-carnitine improves glucose disposal in type 2 diabetic patients. J Am Coll Nutr 1999;18:77–82.
- 12. Alhazmi A, Stojanovski E, McEvoy M, Garg ML. The association between dietary patterns and type 2 diabetes: a systematic review and meta-analysis of cohort studies. J Hum Nutr Diet 2014;27:251–60.
- McEvoy CT, Cardwell CR, Woodside JV, Young IS, Hunter SJ, McKinley MC. A posteriori dietary patterns are related to risk of type 2diabetes: findings from a systematic review and meta-analysis. J Acad Nutr Diet 2014;114:1759–75.
- 14. Salas-Salvadó J, Bulló M, Estruch R, Ros E, Covas MI, Ibarrola-Jurado N, Corella D, Arós F, Gómez-Gracia E, Ruiz-Gutiérrez V, et al. Prevention of diabetes with Mediterranean diets: a subgroup analysis of a randomized trial. Ann Intern Med 2014;160:1–10.
- Cornelis MC, Hu FB. Systems epidemiology: a new direction in nutrition and metabolic disease research. Curr Nutr Rep 2013;2.
- Estruch R, Ros E, Salas-Salvadó J, Covas MI, Corella D, Arós F, Gómez-Gracia E, Ruiz-Gutiérrez V, Fiol M, Lapetra J, et al. Primary prevention of cardiovascular disease with a Mediterranean diet. N Engl J Med 2013;368:1279–90.
- 17. Martínez-González MÁ, Corella D, Salas-Salvadó J, Ros E, Covas MI, Fiol M, Wärnberg J, Arós F, Ruíz-Gutiérrez V, Lamuela-Raventós RM, et al. Cohort profile: design and methods of the PREDIMED study. Int J Epidemiol 2012;41:377–85.
- 18. Wang TJ, Larson MG, Vasan RS, Cheng S, Rhee EP, McCabe E, Lewis GD, Fox CS, Jacques PF, Fernandez C, et al. Metabolite profiles and the risk of developing diabetes. Nat Med 2011;17:448–53.
- Rhee EP, Cheng S, Larson MG, Walford GA, Lewis GD, McCabe E, Yang E, Farrell L, Fox CS, O'Donnell CJ, et al. Lipid profiling identifies a triacylglycerol signature of insulin resistance and improves diabetes prediction in humans. J Clin Invest 2011;121:1402–11.
- Lewis GD, Farrell L, Wood MJ, Martinovic M, Arany Z, Rowe GC, Souza A, Cheng S, McCabe EL, Yang E, et al. Metabolic signatures of exercise in human plasma. Sci Transl Med 2010;2:33ra37.
- Gohil VM, Sheth SA, Nilsson R, Wojtovich AP, Lee JH, Perocchi F, Chen W, Clish CB, Ayata C, Brookes PS, et al. Nutrient-sensitized screening for drugs that shift energy metabolism from mitochondrial respiration to glycolysis. Nat Biotechnol 2010;28:249–55.
- Rhee EP, Souza A, Farrell L, Pollak MR, Lewis GD, Steele DJ, Thadhani R, Clish CB, Greka A, Gerszten RE, et al. Metabolite profiling identifies markers of uremia. J Am Soc Nephrol 2010;21:1041–51.
- Shaham O, Slate NG, Goldberger O, Xu Q, Ramanathan A, Souza AL, Clish CB, Sims KB, Mootha VK. A plasma signature of human mitochondrial disease revealed through metabolic profiling of spent media from cultured muscle cells. Proc Natl Acad Sci USA 2010;107:1571–5.
- Zhong L, D'Urso A, Toiber D, Sebastian C, Henry RE, Vadysirisack DD, Guimaraes A, Marinelli B, Wikstrom JD, Nir T, et al. The histone deacetylase Sirt6 regulates glucose homeostasis via Hif1alpha. Cell 2010;140:280–93.
- American Diabetes Association. Diagnosis and classification of diabetes mellitus. Diabetes Care 2008;31:55–60.
- Elosua R, Marrugat J, Molina L, Pons S, Pujol E; MARATHOM Investigators. Validation of the Minnesota Leisure Time Physical Activity Questionnaire in Spanish men. Am J Epidemiol 1994;139:1197–209.
- Schröder H, Fitó M, Estruch R, Martínez-González MA, Corella D, Salas-Salvadó J, Lamuela-Raventós R, Ros E, Salaverría I, Fiol M, et al. A short screener is valid for assessing Mediterranean diet adherence among older Spanish men and women. J Nutr 2011;141;1140–5.
- Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF TR. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. Diabetologia 1985;28:412–9.
- Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. J R Stat Soc Ser B Methodol 1995;57:289–300.

- Stephens FB, Constantin-Teodosiu D, Greenhaff PL. New insights concerning the role of carnitine in the regulation of fuel metabolism in skeletal muscle. J Physiol 2007;581:431

 –44.
- 31. Schwab U, Törrönen A, Meririnne E, Saarinen M, Alfthan G, Aro A, Uusitupa M. Orally administered betaine has an acute and dose-dependent effect on serum betaine and plasma homocysteine concentrations in healthy humans. J Nutr 2006;136:34–8.
- Huang T, Ren J, Huang J, Li D. Association of homocysteine with type 2 diabetes: a meta-analysis implementing Mendelian randomization approach. BMC Genomics 2013;14:867.
- 33. Cho NH, Lim S, Jang HC, Park HK, Metzger BE. Elevated homocysteine as a risk factor for the development of diabetes in women with a previous history of gestational diabetes mellitus: a 4-year prospective study. Diabetes Care 2005;28:2750–5.
- 34. Lever M, Atkinson W, Slow S, Chambers ST, George PM. Plasma and urine betaine and dimethyl-glycine variation in healthy young male subjects. Clin Biochem 2009;42:706–12.
- 35. Wang Z, Tang WH, Buffa JA, Fu X, Britt EB, Koeth RA, et al. Prognostic value of choline and betaine depends on intestinal microbiota-generated metabolite trimethylamine-N-oxide. Eur Hear J 2014;35:904–10.
- Dambrova M, Latkovskis G, Kuka J, Strele I, Konrade I, Grinberga S, Hartmane D, Pugovics O, Erglis A, Liepinsh E. Diabetes is associated with higher trimethylamine N-oxide plasma levels. Exp Clin Endocrinol Diabetes 2016;124(4):251–6.
- Cho CE, Caudill MA. Trimethylamine-N-oxide: friend, foe, or simply caught in the cross-fire? Trends Endocrinol Metab 2017;28:121–30.
- Trøseid M, Hov JR, Nestvold TK, Thoresen H, Berge RK, Svardal A, Lappegård KT. Major increase in microbiota-dependent proatherogenic metabolite TMAO one year after bariatric surgery. Metab Syndr Relat Disord 2016;14:197–201.
- Shan Z, Sun T, Huang H, Chen S, Chen L, Luo C, Yang W, Yang X, Yao P, Cheng J, et al. Association between microbiota-dependent metabolite trimethylamine-N-oxide and type 2 diabetes. Am J Clin Nutr 2017;106(3):888–94.
- Tang WH, Wang Z, Levison BS, Koeth RA, Britt EB, Fu X, Wu Y, Hazen SL. Intestinal microbial metabolism of phosphatidylcholine and cardiovascular risk. N Engl J Med 2013;368:1575–84.
- 41. McEntyre CJ, Lever M, Chambers ST, George PM, Slow S, Elmslie JL, Florkowski CM, Lunt H, Krebs JD. Variation of betaine, N,N-dimethylglycine, choline, glycerophosphorylcholine, taurine and trimethylamine-N-oxide in the plasma and urine of overweight people with type 2 diabetes over a two-year period. Ann Clin Biochem 2015;52:352–60.
- 42. Wallace M, Morris C, O'Grada CM, Ryan M, Dillon ET, Coleman E, Gibney ER, Gibney MJ, Roche HM, Brennan L. Relationship between the lipidome, inflammatory markers and insulin resistance. Mol Biosyst 2014;10:1586–95.
- 43. Floegel A, Stefan N, Yu Z, Mühlenbruch K, Drogan D, Joost HG, Fritsche A, Häring HU, Hrabě de Angelis M, et al. Identification of serum metabolites associated with risk of type 2 diabetes using a targeted metabolomic approach. Diabetes 2013;62: 639–48.
- 44. Yea K, Kim J, Yoon JH, Kwon T, Kim JH, Lee BD, Lee HJ, Lee SJ, Kim JI, Lee TG, et al. Lysophosphatidylcholine activates adipocyte glucose uptake and lowers blood glucose levels in murine models of diabetes. J Biol Chem 2009;284:33833–40.
- 45. Nishimukai M, Maeba R, Yamazaki Y, Nezu T, Sakurai T, Takahashi Y, Hui SP, Chiba H, Okazaki T, Hara H. Serum choline plasmalogens, particularly those with oleic acid in sn-2, are associated with proatherogenic state. J Lipid Res 2014;55:956–65.
- Pacifico L, Nobili V, Anania C, Verdecchia P, Chiesa C. Pediatric nonalcoholic fatty liver disease, metabolic syndrome and cardiovascular risk. World J Gastroenterol 2011;17:3082–91.
- Puri P, Wiest MM, Cheung O. The plasma lipidomic signature of nonalcoholic steatohepatitis. Hepatology 2009;50: 1827–38.
- Amenta F, Tayebati SS. Pathways of acetylcholine synthesis, transport and release as targets for treatment of adult-onset cognitive dysfunction. Curr Med Chem 2008;15:488–98.
- Merlin J, Evans BA, Csikasz RI, Bengtsson T, Summers RJ, Hutchinson DS. The M3-muscarinic acetylcholine receptor stimulates glucose uptake in L6 skeletal muscle cells by a CaMKK-AMPK-dependent mechanism. Cell Signal 2010;22:1104–13.